

Investigation of Host–Guest Inclusion Complexes of a Neurotransmitter inside into α - and β -Cyclodextrins through Hydrophobic and Hydrophilic Interactions by Physicochemical Approach

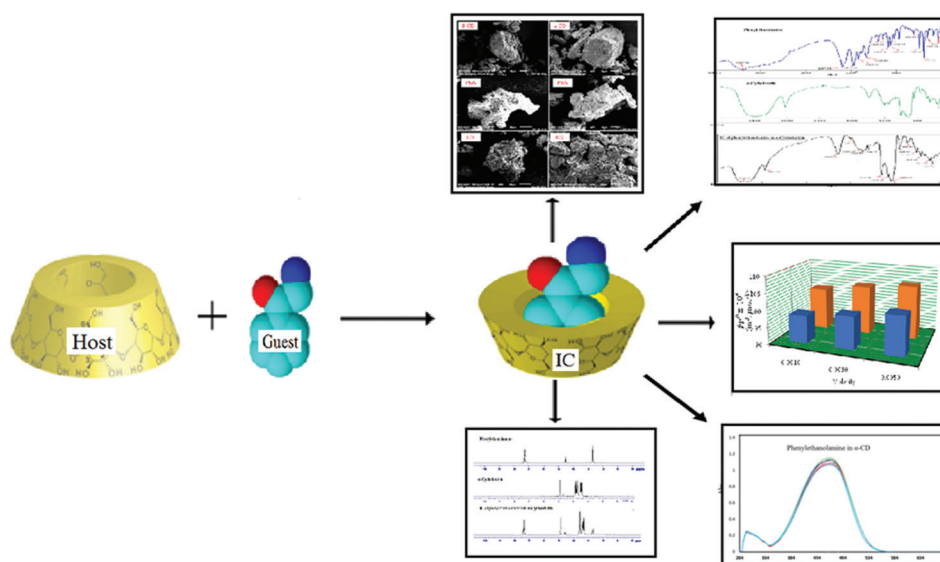
Habibur Rahaman, Kalipada Sarkar, Mitali Kundu, Mahendra Nath Roy*

Department of Chemistry, Pedong Government College, Kalimpong, West Bengal, 734311, India

ABSTRACT

In the present work, the formation of inclusion complex of a neurotransmitter, phenylethanolamine with α -cyclodextrin (α -CD) and β -CD (β -CD) was examined from volumetric, viscometric, and spectroscopic studies. The supramolecular interaction between CD and phenylethanolamine has been characterized by volumetric and viscometric and spectral studies. However, the stoichiometry of host–guest of the inclusion complexes was ascertained from Job's plot calculation from ultraviolet–visible spectroscopy. The association/binding constant of the formation of inclusion complex was calculated from Benesi–Hildebrand equation. The infrared (IR) and $^1\text{H-NMR}$ spectroscopy also support the formation of inclusion complexes and the probable manner of inclusion was defined from $^1\text{H-NMR}$ and two-dimensional Roesy NMR spectroscopies.

Key words: Phenylethanolamine, Job's plot, cyclodextrins, Benesi–Hildebrand equation.



Graphical Abstract

1. INTRODUCTION

Phenylethanolamine is known as β -hydroxyphenethylamine is a neurotransmitter which is structurally similar to others trace amine such as norepinephrine, dopamine, and epinephrine [1,2].

It is endogenous chemical which transmits signal across a chemical synapse from one neuron to another neuron. The neurotransmitters produced from synaptic vesicles are received by specific neurotransmitter receptors on the cell membrane of the postsynaptic neuron as shown in Scheme 1. This neuron may be connected to many more neurons and pass the information to adjacent neuron [3-5].

The neurotransmitters are clinically used for the treatment of several neurological and psychiatric disorders such as schizophrenia, Parkinson's disease, bipolar disorder, attention deficit, Huntington's disease, and hyperactivity disorder. It regulates the blood pressure, respiration, and body

*Corresponding author:

E-mail: mahendraroy2002@yahoo.co.in

ISSN NO: 2320-0898 (p); 2320-0928 (e)

DOI: 10.22607/IJACS.2019.703005

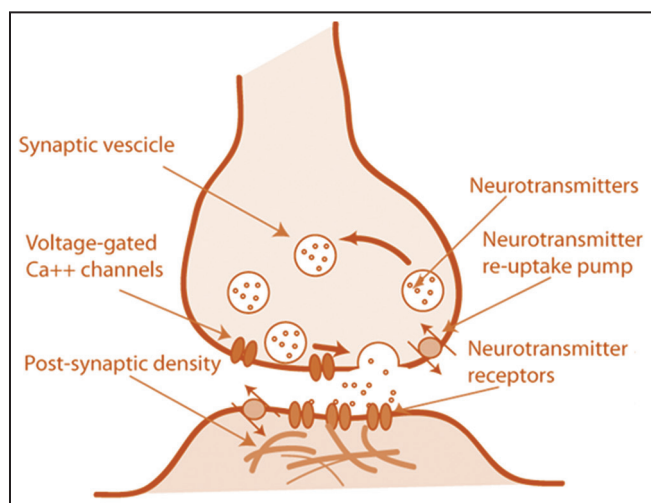
Received: 19th February 2019;

Revised: 11th July 2019;

Accepted: 21th July 2019

temperature, the secretion of hormones from the pituitary gland, the regulation of α_2 -adrenoceptors in the hypothalamus, etc. Phenylethanolamine is responsible for the production of enzyme phenylethanolamine-N-methyltransferase for the conversion of norepinephrine into epinephrine [6-8]. Phenylethanolamine-N-methyltransferase catalyzes the biosynthesis of epinephrine from norepinephrine by transferring methyl group from S-adenosyl-l-methionine [9].

The cyclodextrins (CDs) are the cyclic oligosaccharides of glucopyranose units. There are three kinds of CDs, α -CD (α -CD), β -CD (β -CD), and γ -CD (γ -CD) containing 6, 7, and 8 glucopyranose



Scheme 1: Process of transmission of signals from one neuron to another neuron.

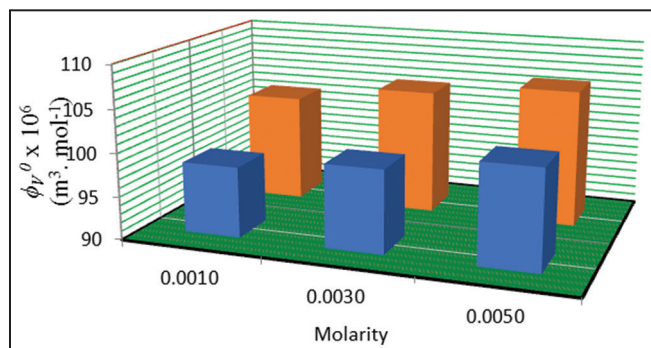


Figure 1: Variation of limiting apparent molar volumes (ϕ_v^0) of phenylethanolamine in aqueous α - and β -CDs solution with molarity (red and blue color for α - and β -CDs, respectively).

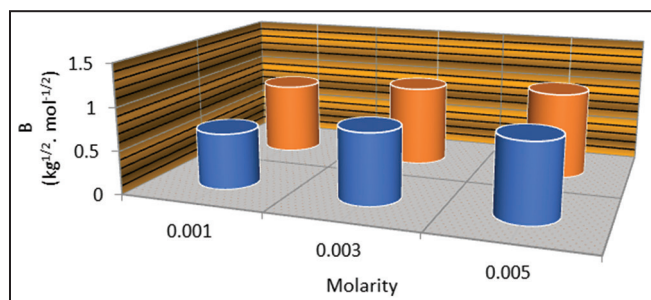


Figure 2: Variation of viscosity B coefficient of phenylethanolamine in aqueous α - and β -CDs solution with molarity (red and blue column for α - and β -CDs, respectively).

units, respectively, connected with α -(1-4) linkages [10,11]. The two-dimensional (2D) and three-dimensional structures of CDs are shown in Schemes 2 and 3. The special characteristic of CD is that its inner cavity is hydrophobic in nature, whereas exterior side is hydrophilic in nature. This kind of exceptional property allows CD for complexation with diverse molecules such as vitamins, amino acids, drugs, ionic liquids, hormones, polymers, and dyes [12-16]. The H3 and H5 protons of CDs are located inside the cavity and the H1, H2, and H4 protons are located outside the cavity as shown in Scheme 3. Another significant feature of CD is that the H3 proton is situated close to the wider rim and H5 proton is situated close to the narrower rim of the CD. The hydrophobic or alkyl parts of the compound enter inside the hydrophobic cavity of CDs and thus forming a stable inclusion complex [17] and the hydrophilic or ionic part of the compound exposes outside the cavity of CDs, thus forming an inclusion complex. The complexation of any compound inside CD increases the solubility, stability against heat, light, oxidation, and bioavailability and reduces volatility of the encapsulated molecules without disturbing its structure.

In our present work, we studied the formation of inclusion complex of phenylethanolamine inside the cavity of α - and β -CDs. Various physicochemical parameters from volumetric and viscometric studies and spectrometric methods were used to inspect the inclusion phenomenon. The inclusion complexes so formed may be used medically for its better performance as drugs.

2. EXPERIMENTAL SECTION

2.1. Materials

The neurotransmitter, phenylethanolamine was procured from TCI Chemicals (Japan) Pvt. Ltd., and α - and β -CDs were procured from Sigma-Aldrich, Germany. All these chemicals were used as purchased as their mass fraction purity was >0.98 .

2.2. Apparatus and Procedure

The solutions were prepared with triply distilled water. The weight was measured with Mettler AG-285 electronic balance having precession $\pm 0.0003 \times 10^{-3}$ kg.

Anton Paar Density Meter (DMA 4500M) with a precision of 0.00001×10^{-3} (kg/m^3) was employed to measure the density (ρ) of the solutions at different temperatures. The calibration of the density meter was done using doubly distilled water and dry air.

The viscosities of the solutions were taken with Brookfield DV-III Ultra Programmable Rheometer with spindle size-42. The viscometer was connected with Brookfield Digital Bath TC-500. The machine was calibrated with doubly distilled water and purified methanol at 298.15 K before recording the viscosities of our studied solutions. The uncertainty in viscosity is within ± 0.003 mPa/s.

Ultraviolet (UV)-visible absorption spectra of phenylethanolamine solution with successive addition of CDs were taken at 298.15 K by JASCO V-530 UV-visible spectrophotometer. In our present work, a probe methyl orange (MO) was used since the studied neurotransmitter does not absorb in the UV-visible range.

The Fourier-transform infrared (FT-IR) spectra were recorded with PerkinElmer FT-IR spectrometer after preparing the KBr disk of phenylethanolamine, CDs, and inclusion complexes of it. The KBr disk is prepared by mixing 100 mg of carefully dried pure KBr and 1 mg of the compound to be studied.

$^1\text{H-NMR}$ and NMR-ROSEY spectra were taken at 298 K in D_2O by Bruker Avance 400 MHz spectrometer.

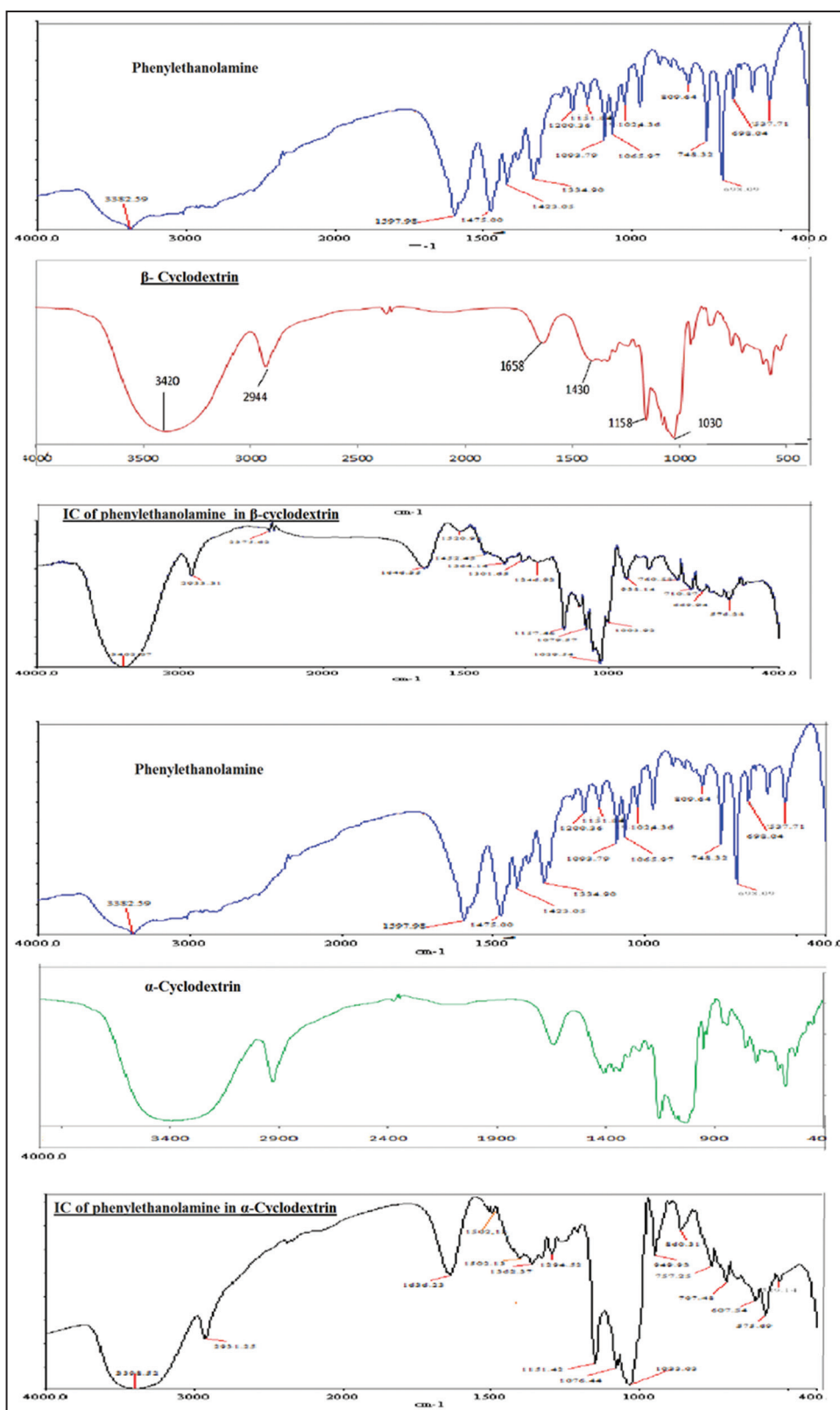


Figure 3: FTIR spectra of phenylethanolamine and inclusion complexes of it in α - and β -CD at 298.15 K.

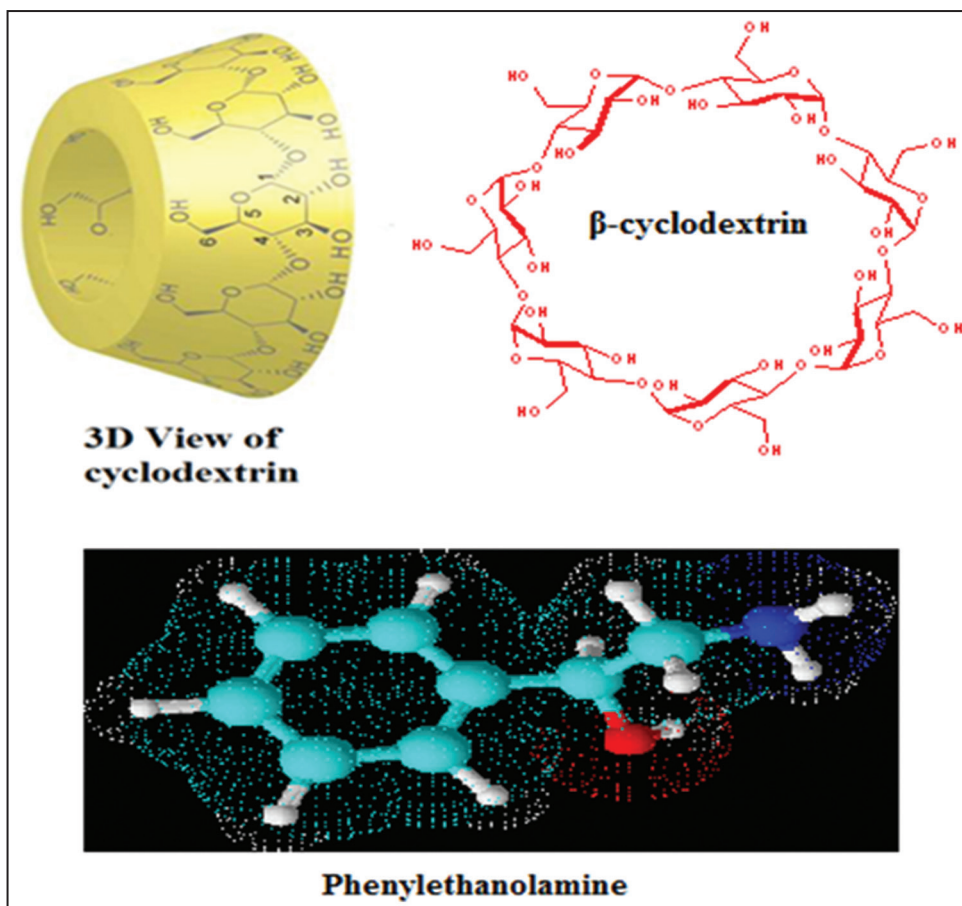
3. RESULTS AND DISCUSSION

3.1. Density Study

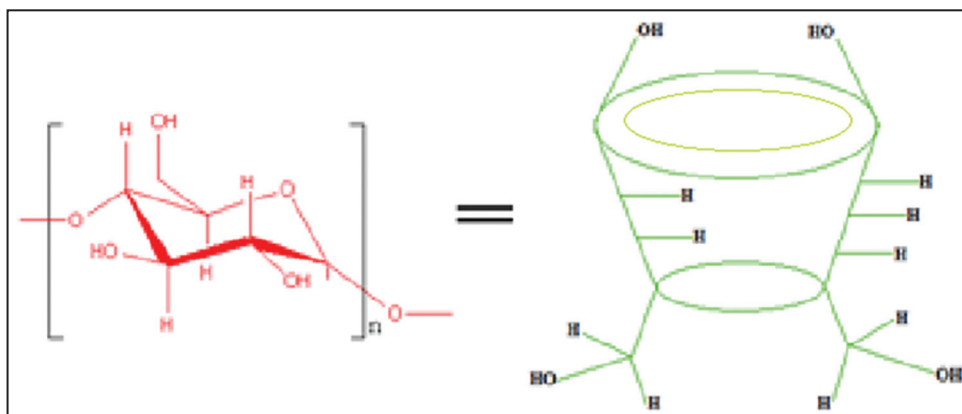
We may get valuable information about the interactions between phenylethanolamine and CD molecules from volumetric study. The values of densities (ρ) and viscosities (η) of different molar aqueous

solutions of phenylethanolamine in α - and β -CDs at 298.15 K are shown in Tables S1 and S2, respectively. We calculated the apparent molar volume, ϕ_v from densities of different molarities of phenylethanolamine in aqueous solution of α - and β -CDs of varying molarities from the following equation [18].

$$\phi_v = M / \rho - 1000 (\rho - \rho_0) / (m \rho \rho_0) \quad (1)$$



Scheme 2: Structure of cyclodextrin and phenylethanolamine.



Scheme 3: Structure of cyclodextrins.

The limiting apparent molar volume, ϕ_V^0 was evaluated by least square treatment of the plots of ϕ_V against \sqrt{m} using the renowned Masson equation [19].

$$\phi_V = \phi_V^0 + S_V^* \sqrt{m} \quad (2)$$

The ϕ_V^0 provides the information about solute-solvent interaction. The variations of limiting apparent molar volumes (ϕ_V^0) of phenylethanolamine are shown in Table S3 and Figure 1. It is noticed that ϕ_V^0 values increase with increase in molarity of phenylethanolamine and CDs which specifies that solute and cosolute interactions increase with increasing concentration of phenylethanolamine and CDs are shown in Table S3 and Figure 1. The interaction is assumed to be arisen from the H-bonding between $-NH_2$ and $-OH$ groups of

phenylethanolamine and $-OH$ group of CDs. The higher ϕ_V^0 value of phenylethanolamine in α -CD than β -CD indicates that the former interacts strongly with phenylethanolamine than later. This cannot be explained with H-bonding only. If the interaction was arisen only from H-bonding, then ϕ_V^0 values for both α -CD and β -CD should be same. The difference in ϕ_V^0 values of phenylethanolamine in α -CD and β -CD can be explained on the basis of formation of inclusion complex which occurs due to another kind of non-bonding interaction known as hydrophobic-hydrophobic interaction. The hydrophobic aryl part of phenylethanolamine enters inside the hydrophobic interior of CD forming inclusion complex and exert hydrophobic-hydrophobic interaction [20]. The higher ϕ_V^0 values of phenylethanolamine in α -CD than β -CD may be explained from their cavity sizes. We know that the

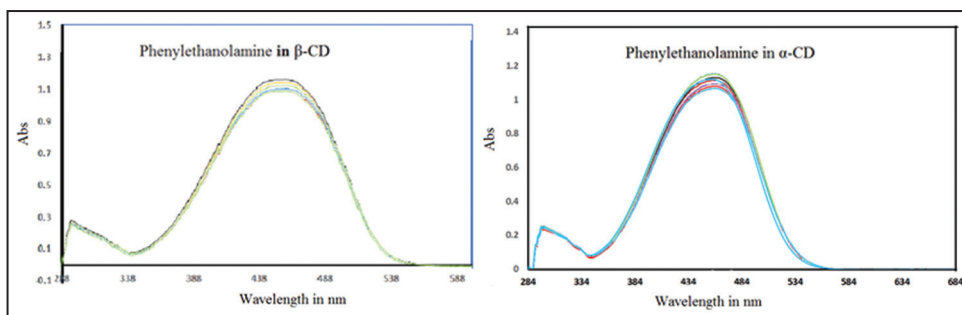


Figure 4: Ultraviolet-visible spectra of phenylethanolamine in different concentrations of CDs using MO as probe.

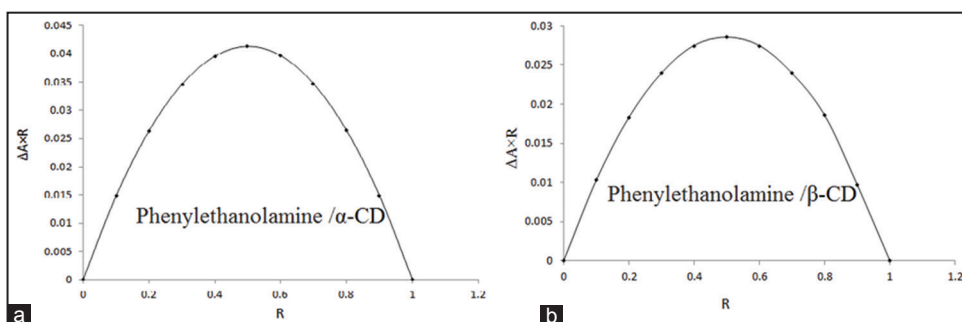


Figure 5: Job's plot of different concentrations of (a) β -CD in phenylethanolamine (b) α -CD in phenylethanolamine using MO as probe.

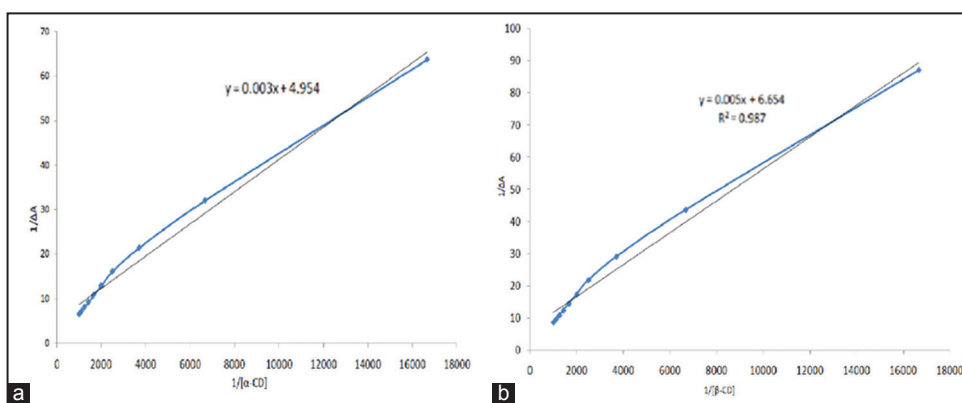


Figure 6: (a) Plot of $1/\Delta A$ against $1/[\alpha\text{-CD}]$ for examining stoichiometry of inclusion complexes. (b) Plot of $1/\Delta A$ against $1/[\beta\text{-CD}]$ for examining stoichiometry of inclusion complexes.

cavity size of α - and β -CD is 4.7–5.3 Å and 6.0–6.5 Å, respectively. The smaller cavity size of α -CD provides a better situation for hydrophobic-hydrophobic interaction with phenylethanolamine than β -CD which has comparatively larger cavity size. For this reason, ϕ_v^0 of phenylethanolamine is higher in α -CD than in β -CD. Hence, volumetric study gives us an idea about the formation of inclusion of phenylethanolamine with CDs.

3.2. Viscosity Study

Viscosity study is also helpful for inspecting the inclusion complex of phenylethanolamine with CDs [21]. From the viscosities of the phenylethanolamine in different molarities of aqueous CDs solution, we can calculate the viscosity A and B coefficient using famous Jones-Dole equation given below.

$$(\eta/\eta_0 - 1)/\sqrt{m} = A + B \sqrt{m} \quad (3)$$

Where, η_0 and η are the viscosities of the solvent (aqueous solution of cosolute) and solution, respectively. The viscosity coefficients A

and B values are obtained from intercept and slope of the straight line obtained from the plot of $(\eta/\eta_0 - 1)/\sqrt{m}$ against \sqrt{m} and reported in Table S3 and Figure 2. The viscosity B coefficient gives the information about the solute-solvent interaction [22,23]. The values of viscosity B coefficients of phenylethanolamine increase with increasing concentration of phenylethanolamine and CDs. This indicates that interaction between phenylethanolamine and the CD increases with increasing concentration of CD. The interaction arises from H-hydrogen bonding between -OH group of CD and -OH and -NH₂ groups of phenylethanolamine. It is observed that B values for phenylethanolamine/ α -CD and phenylethanolamine/ β -CD systems are different which cannot be explained by H-bonding only. Similar kind of observation was obtained in volumetric study also which may be explained by hydrophobic-hydrophobic interaction due to the formation of inclusion complex. The hydrophobic aryl part of phenylethanolamine enters into the hydrophobic interior of CD and forms inclusion complex by hydrophobic-hydrophobic interaction. The higher B value for phenylethanolamine/ α -CD system

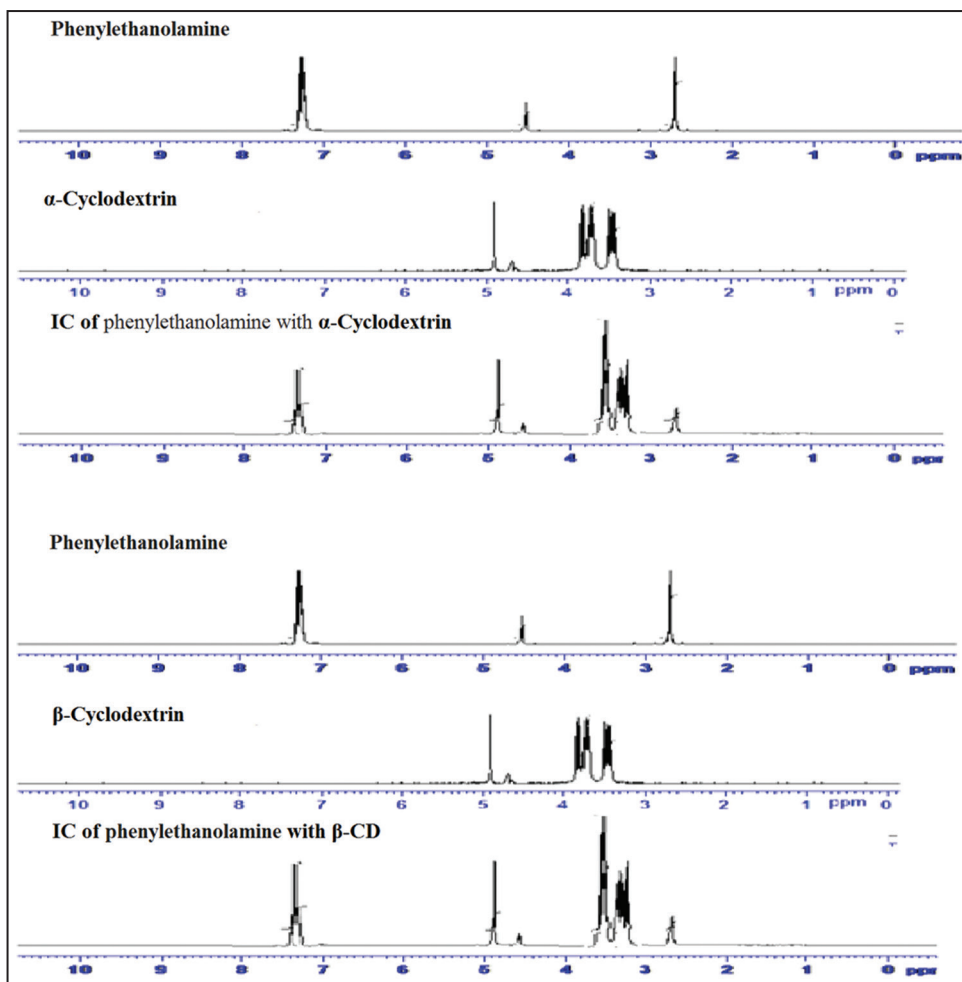


Figure 7: $^1\text{H-NMR}$ spectra of phenylethanolamine and inclusion complexes of it with α - and β -CDs in D_2O at 299.15 K.

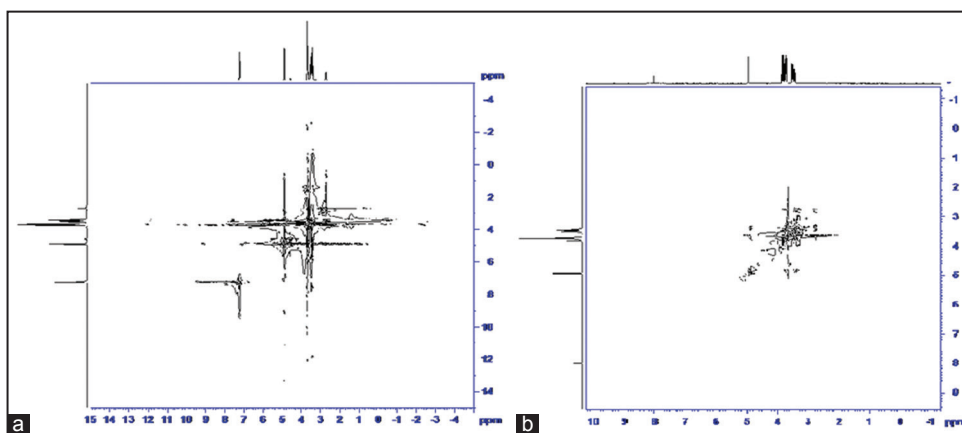


Figure 8: NMR Roesy spectra of inclusion complexes of (a) phenylethanolamine and α -CD (b) phenylethanolamine and β -CD.

than phenylethanolamine/ β -CD systems indicates that α -CD is more favorable in forming inclusion complex than β -CD [24]. The smaller cavity size of α -CD provides a better situation for encapsulation than β -CD.

3.3. FT-IR Spectroscopy

The FT-IR study is a very reliable technique for investigation the inclusion phenomena [25-27]. The FT-IR spectra of pure

phenylethanolamine, α - and β -CD CDs, and their inclusion complexes are given in Figure 3.

Some characteristic frequencies of phenylethanolamine, α - and β -CD CDs, and their inclusion complexes are given below. The frequencies for $-\text{OH}$ group of both the α - and β -CD shifted to the lower frequencies which may be considered due to the presence of H-bonding between $-\text{OH}$ groups of the CDs and $-\text{OH}$ and $-\text{NH}_2$ groups of phenylethanolamine. The peaks positions for aryl part of phenylethanolamine and hydrogen

present in the interior of CDs changed due to encapsulation of aryl group into the cavity of CD.

The frequencies of different groups are as follows:

Phenylethanolamine: 3458 cm^{-1} (O-H stretch), 2813.55 cm^{-1} (C-H stretch for $-\text{CH}_2$), 1597.98 cm^{-1} ($-\text{C}=\text{C}$ stretch for aromatic), 1589 cm^{-1} (C=C), 1475 cm^{-1} (C-C stretch for aromatic), and 1423.05 cm^{-1} (Bending- CH_2), α -CD: 3434 cm^{-1} (stretching of O-H), 2930 cm^{-1} (stretching of $-\text{CH}$ from $-\text{CH}_2$), 1420 cm^{-1} (bending $-\text{CH}$), 1160 cm^{-1} (bending of C-O-C), 1080 cm^{-1} (stretching of C-C-O), and 956 cm^{-1} (vibration α -1,4 linkage).

β -CD: 3327 cm^{-1} (stretching of O-H), 2944 cm^{-1} (stretching of $-\text{CH}$ from $-\text{CH}_2$), 1430 cm^{-1} (bending $-\text{CH}$), 1158 cm^{-1} (bending of C-O-C), 1030 cm^{-1} (stretching of C-C-O), and 953 cm^{-1} (vibration α -1,4 linkage).

Phenylethanolamine/ α -CD inclusion complex: 3398.52 cm^{-1} (stretching of O-H of α -CD), 2931 cm^{-1} (stretching of $-\text{C-H}$), 1456.02 cm^{-1} (bending of $-\text{C-H}$), and 1502.13 cm^{-1} (C-C stretch for aromatic).

Phenylethanolamine/ β -CD inclusion complex: 3402.02 cm^{-1} (stretching of O-H of β -CD), 2922.31 cm^{-1} (stretching of $-\text{C-H}$), 1520 cm^{-1} (C-C stretch for aromatic), and 1452.45 cm^{-1} (bending of $-\text{C-H}$).

3.4. UV-Visible Spectroscopy Investigation

A significant indication about the formation inclusion complex may be obtained from UV-visible spectroscopy [28]. Since our studied compounds phenylethanolamine and CDs do not absorb in the UV-visible range, we used MO as probe.

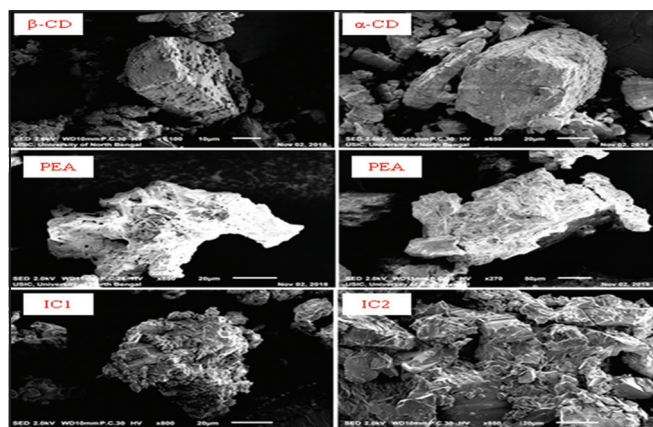


Figure 9: SEM images of α -CD, β -CD, and their inclusion complexes with phenylethanolamine.

The absorption spectra of phenylethanolamine in different molarities of CDs were recorded and shown in Figure 4.

The stoichiometry of host and guest of inclusion complex may be obtained from absorbance values by plotting the graph of $\Delta A \times R$ against R which is known as Job's plot [29-31] [Figure 5].

Where, R is concentration ratio

$R = [\text{PEA}]/([\text{PEA}]+[\text{CD}])$ and ΔA represents the difference in absorbance of phenylethanolamine with and without CD at 298.15 K. R values at the maxima of Job's plot indicate the stoichiometry of host and guest of inclusion complexes.

$R = 0.5, 0.33,$ and 0.66 at the maxima indicate the 1:1, 1:2, and 2:1 host-guest stoichiometry of the inclusion complexes.

R value in case of phenylethanolamine/CD system is 0.5 which indicates the 1:1 stoichiometry.

The association constant of inclusion complex formation (Schemes 4 and 5) for phenylethanolamine/CD system, K_a may be determined from the absorptivity values of UV-visible spectra. We used the well-known Benesi-Hildebrand equation to estimate the association constants (K_a) of inclusion complex formation [32].

$$\frac{1}{\Delta A} = \frac{1}{\Delta \epsilon K [\text{Guest}]} \times \frac{1}{[\text{Host}]} + \frac{1}{\Delta \epsilon}$$

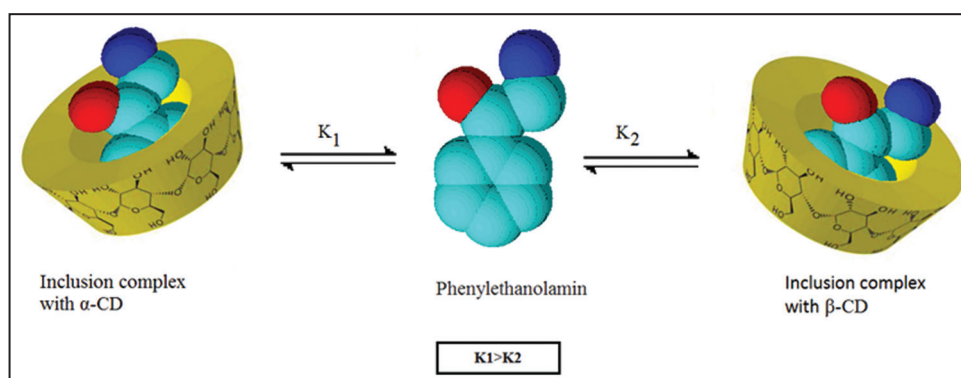
The spectra of phenylethanolamine at constant molarity were recorded with varying concentration of CD in the presence of the probe (MO).

Where, ΔA denotes the difference in absorbance of phenylethanolamine in the presence and absence of CDs, $[\text{Guest}]$ and $[\text{Host}]$ represent the concentration of phenylethanolamine and CD, respectively, and $\Delta \epsilon$ denotes the molar absorption coefficient difference in the presence and absence of CDs. The plot of $1/\Delta A$ against $1/[\text{CDs}]$ gives a straight line with an

intercept $1/\Delta \epsilon$ and a slope of $\frac{1}{\Delta \epsilon K [\text{Guest}]}$. The association constant, K_a may be achieved by dividing the intercept with the slope of the plot at a certain concentration of ionic liquid. The K_a evaluated from Benesi-Hildebrand equation for phenylethanolamine/ α -CD system is $1.216 \times 10^4 \text{ M}^{-1}$ and for phenylethanolamine/ β -CD system is $1.331 \times 10^4 \text{ M}^{-1}$.

3.5. $^1\text{H-NMR}$ Study

$^1\text{H-NMR}$ study gives us important information about the formation of host-guest inclusion complex [33]. The $^1\text{H-NMR}$ spectra of phenylethanolamine, CDs, and their IC were recorded in D_2O at 298.15 K and are shown in Figure 7. Remarkable chemical shifts of various protons of the inclusion complexes from host and guest molecules noted. It is known that H3 and H5 protons of CDs are positioned inside the

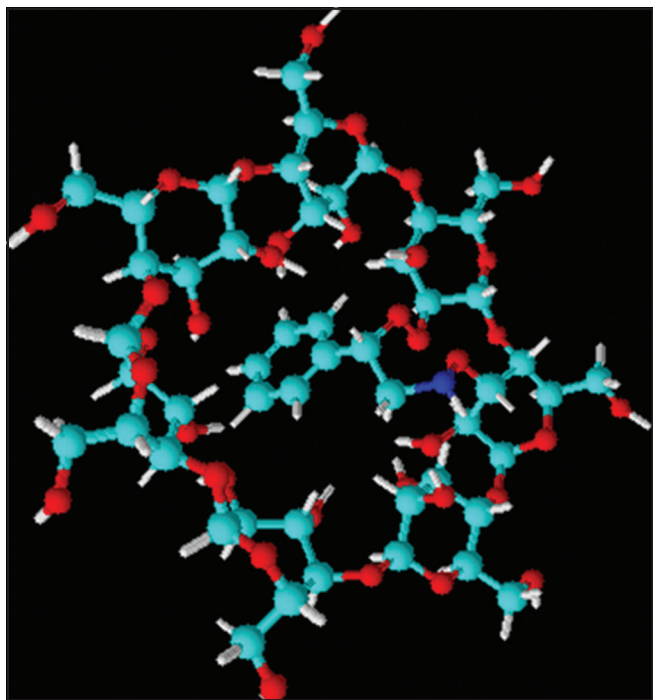


Scheme 4: Formation of inclusion complexes of phenylethanolamine with α - and β -CDs, respectively, and the order of association constants.

cavity, whereas the H1, H2, and H4 protons are positioned outside the cavity [34,35]. Another important feature of CD is that the H3 proton is situated closer to the wide rim and H5 proton is situated closer to the narrow rim of the CD. The alkyl part of phenylethanolamine is assumed to be held inside CD cavity through hydrophobic-hydrophobic interaction without forming or breaking any bond. Due to the encapsulation of aryl part of the phenylethanolamine inside the cavity of CDs, notable upfield chemical shift of the H3 and H5 protons of CDs and downfield chemical shift of protons of aryl part of the ionic liquid took place [33]. Comparatively larger chemical shift for H3 proton than the H5 proton supports that the encapsulation of aryl part of phenylethanolamine inside CD molecules occurs through the wider rim of CD as shown in Scheme 6.

3.5.1. $^1\text{H-NMR}$ data

Phenylethanolamine: [$^1\text{H-NMR}$ (300 MHz, D_2O)]: δ 2.694–2.732 (2H, m), 4.527–4.559 (1H, t), 7.210–7.315 (5H, m).



Scheme 5: Diagrammatic representation showing probable geometrical configurations of inclusion complexes of phenylethanolamine with α - and β -CDs.

α -CD: [$^1\text{H-NMR}$ (300 MHz, D_2O)]: δ 3.42–3.43 (1H, $j=9.00$ Hz), 3.51–3.52 (1H, $j=10$ Hz), 3.74–3.83 (1H, m), 3.87–3.91 (1H, $J=9$ Hz), 4.96–4.97 (1H, $J=3$ Hz), 7.44–7.535 (6H, d).

β -CD: [$^1\text{H-NMR}$ (300 MHz, D_2O)]: δ 3.41–3.42 (1H, $j=9.00$ Hz), 3.53–3.56 (1H, $j=10$ Hz), 3.75–3.77 (2H, m), 3.82–3.83 (1H, $J=9$ Hz), 4.97–4.98 (1H, $J=3$ Hz).

Phenylethanolamine/ β -CD: [$^1\text{H-NMR}$ (300 MHz, D_2O)]: δ 2.684–2.711 (2H, m), 3.267–3.275 (1H, m), (1H, m), 3.292–3.298 (1H, m), 3.558–3.567 (1H, m), 3.732–3.748 (1H, m), 4.97–4.98 (1H, d), 7.46–7.543 (6H, d).

Phenylethanolamine/ α -CD: [$^1\text{H-NMR}$ (300 MHz, D_2O)]: δ 2.688–2.716 (2H, m), 3.265–3.272 (1H, m), (1H, m), 3.288–3.292 (1H, m), 3.552–3.563 (1H, m), 3.725–3.774 (1H, m), 4.96–4.97 (1H, d), 7.44–7.535 (6H, d).

3.6. 2D Roesy NMR

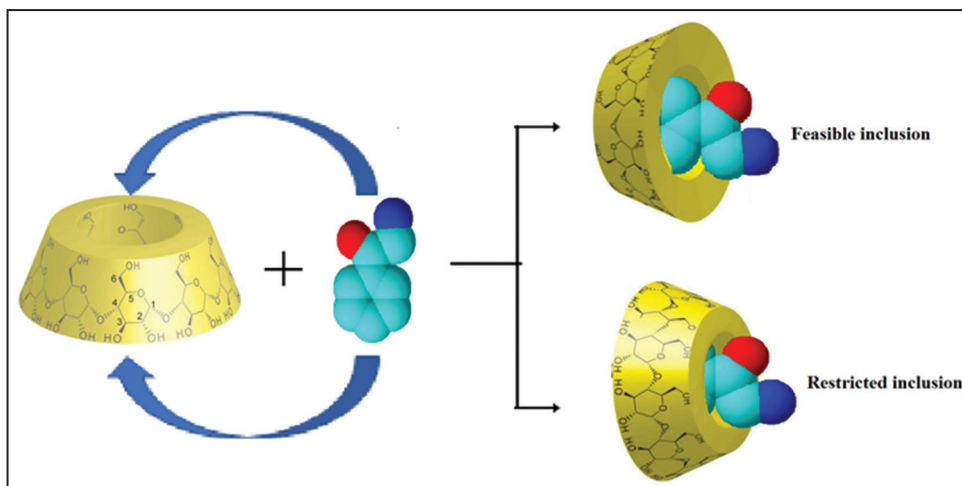
2D ROESY NMR study is another vital technique to investigate the formation of inclusion complexes [36,37]. We are aware that two neighboring protons placed within a distance of 0.4 nm can exert a nuclear Overhauser effect which may be established by 2D ROESY NMR (rotating frame NOE spectroscopy). The ROESY NMR spectra of inclusion complexes of phenylethanolamine inside CDs are displayed in Figure 8. The H3 and H5 protons of CDs situated inside the cavity experience nuclear Overhauser effect with the protons of aryl part of phenylethanolamine showing cross-peaks in the ROESY NMR spectra of inclusion complexes.

3.7. Scanning Electron Microscope (SEM)

SEM is an exceptionally familiar method for examining the surface texture and particle size of solid materials [38-41]. From the SEM images of the drug molecule, α -CD, β -CD, and their complexes, it is clear that the SEM images of pure compounds and their Inclusion complexes (ICs) are totally different. Significant morphological changes and changes of surface structures of pure compounds and their ICs are observed. Changes of SEM images of pure hosts and guest molecule indicate about the formation of inclusion complexes.

4. CONCLUSION

The formation of inclusion complexes of phenylethanolamine inside the cavity of α - and β -CDs has been proved by various physicochemical and spectroscopic studies. The volumetric and viscometric studies reveal the existence of considerable interaction between phenylethanolamine



Scheme 6: Feasible and restricted inclusion complex formation of phenylethanolamine with CD.

and CD in aqueous medium. The UV-visible, $^1\text{H-NMR}$, 2D ROESY NMR, and IR spectroscopy firmly establish the formation of inclusion complexes of phenylethanolamine with both CDs. The Job's plot from UV-visible spectroscopy reveals that the inclusion is of 1:1 stoichiometry; however, $^1\text{H-NMR}$ spectroscopy reveals that the encapsulation of aryl part of phenylethanolamine inside the cavity of CDs occurs through the wider rim of CDs.

5. ACKNOWLEDGMENT

Prof. M.N. Roy is indebted to the UGC, New Delhi, India, for giving him 1 time Grant-in-Aid [No.F4-10/2010 (BSR)] to develop research facilities for the scholars. We express our gratefulness to the SAIF, North Eastern Hill University, Shillong, India, for performing the NMR analysis.

6. REFERENCES

1. W. H. Hartung, (1945) Beta-phenethylamine derivatives, *Industrial and Engineering Chemistry*, **37**: 126-136.
2. J. Axelrod, (1966) Methylation reactions in the formation and metabolism of catecholamines and other biogenic amines, *Pharmacological Review*, **18**: 95-113.
3. L. J. Elias, D. M. Saucier, (2005) *Neuropsychology: Clinical and Experimental Foundations*. Boston: Pearson, Boston, MA: Pearson.
4. E. R. Kandel, J. H. Schwartz, T. M. Jessell, editors (2000) *Transmitter Release. Principles of Neural Science*, 4th ed. New York: McGraw-Hill.
5. T.M. Jessell, (2000) In: K. Eric, J. Schwartz, T. Jessell, editros. *Principles of Neural Science*, New York: McGraw-Hill.
6. M. L. Tainter, (1929) Pharmacological actions of phenylethanolamine, *Journal Pharmacology and Experimental Therapeutics*, **36**: 29-54.
7. H. E. Shannon, E. J. Cone, D. Yousefnejad (1981) Physiologic effects and plasma kinetics of phenylethanolamine and its N-methyl homolog in the dog, *Journal of Pharmacology and Experimental Therapeutics*, **217**: 379-385.
8. C. Carpené, J. Galitzky, E. Fontana, C. Atgié, M. Lafontan, M. Berlan, (1999) Selective activation of β_3 -adrenoceptors by octopamine: Comparative studies in mammalian fat cells, *Naunyn-Schmiedebergs Arch. Pharmacology*, **359**: 310-321.
9. G. L. Grunewald, M. R. Seim, R. C. Regier, K. R. Criscione, (2007) Exploring the active site of phenylethanolamine N-Methyltransferase with 1,2,3,4-tetrahydrobenz[h]isoquinoline inhibitors, *Bioorganic and Medicinal Chemistry*, **15**(3): 1298-1310.
10. E. Leyva, E. Moctezuma, J. Strouse, M. A. García-Garibay, (2001) Spectrometric and 2D NMR studies on the complexation of chlorophenols with cyclodextrins, *Journal of Inclusion Phenomena and Macrocyclic Chemistry*, **39**: 41-46.
11. Y. Kawaguchi, T. Nishiyama, M. Okada, M. Kamachi, A. Harada, (2000) Complex Formation of poly (ϵ -caprolactone) with cyclodextrins, *Macromolecules*, **33**: 4472-4477.
12. M. E. Brewster, T. Loftsson, (2007) Cyclodextrins as pharmaceutical solubilizers, *Advanced Drug Delivery Reviews*, **59**: 645-666.
13. A. Harada, T. Nishiyama, Y. Kawaguchi, M. Okada, M. Kamachi, (1997) Preparation and characterization of inclusion complexes of aliphatic polyesters with cyclodextrins, *Macromolecules*, **30**: 7115-7118.
14. S. S. Satav, R. N. Karmalkar, M. G. Kulkarni, N. Mulpuri, G. N. Sastry, (2006) Formation of linear polymers with pendant vinyl groups via inclusion complex mediated polymerization of divinyl monomers, *Journal of the American Chemical Society*, **128**: 7752-7753.
15. S. H. Cho, S. Y. Kim, S. I. Lee, Y. M. Lee, (2006) Hydroxypropyl- β -cyclodextrin inclusion complexes for transdermal delivery: Preparation, inclusion properties, stability and release behaviour, *Journal of Industrial and Engineering Chemistry*, **12**: 50-59.
16. X. Zhu, J. Sun, J. Wu, (2007) Spectrofluorimetric determination of trace Co(II) in aqueous samples with quinalizarin- β -cyclodextrin inclusion complex, *Talanta*, **72**: 237-242.
17. H. Jiao, S. Goh, S. Valiyaveetil, (2002) Inclusion complexes of poly(4-vinylpyridine)-dodecylbenzenesulfonic acid complex and cyclodextrins, *Macromolecules*, **35**: 3997-4002.
18. D. Ekka, M. N. Roy, (2014) Quantitative and qualitative analysis of ionic solvation of individual ions of imidazolium based ionic liquids in significant solution systems by conductance and FT-IR spectroscopy, *RSC Advances*, **4**: 19831-19845.
19. D. O. Masson, (1929) Solute molecular volumes in relation to the solvation and ionization, *Philosophical Magazine*, **8**: 218-223.
20. H. Shekaari, F. Jebali, (2001) Volumetric and conductometric studies of some amino acids in aqueous ionic liquid, 1-hexyl-3-methylimidazolium chloride solutions at 298.15K, *Physics and Chemistry of Liquids*, **49**: 572-587.
21. A. Roy, S. Saha, M. N. Roy, (2016) Study to explore host-guest inclusion complexes of cyclodextrins with biologically active molecules in aqueous environment, *Fluid Phase Equilibria*, **425**: 252-258.
22. R. H. Stokes, R. Mills, (1965) *Viscosity of Electrolytes and Related Properties, International Encyclopedia of Physical Chemistry and Chemical Physics*, London: Pergamon Press.
23. H. J. V. Tyrrell, M. Kennerley, (1968) Viscosity B -coefficients between 5 and 20 for glycolamide, glycine, and N -methylated glycines in aqueous solution, *Journal of the Chemical Society A*, **0**: 2724-2728.
24. M.N. Roy, M.C. Roy, K. Roy, (2015) Investigation of an inclusion complex formed by ionic liquid and β -cyclodextrin through hydrophilic and hydrophobic interactions, *RSC Advances*, **5**: 56717-56723.
25. W. Zhang, X. Gong, Y. Cai, C. Zhang, X. Yu, J. Fan, G. Diao, (2013) Investigation of water-soluble inclusion complex of hypericin with β -cyclodextrin polymer, *Carbohydrate Polymers*, **95**: 366-370.
26. S. Milovanovic, D. Markovic, K. Aksentijevic, D. B. Stojanovic, J. Ivanovic, I. Zizovic, (2016) Application of cellulose acetate for controlled release of thymol, *Carbohydrate Polymers*, **147**: 344-353.
27. J. S. Negi, S. Singh, (2013) Spectroscopic investigation on the inclusion complex formation between amisulpride and γ -cyclodextrin, *Carbohydrate Polymers*, **92**: 1835-1843.
28. Y. Sueishi, T. Ide, (2004) Inclusion complexation of methyl orange with β -cyclodextrin in a roomtemperature ionic liquid, *Zeitschrift für Physikalische Chemie*, **218**: 829-835.
29. P. Job, (1928) Formation and stability of inorganic complexes in solution, *Annali di Chimica Applicata*, **9**: 113-203.
30. J. S. Renny, L. L. Tomasevich, E. H. Tallmadge, D. B. Collum, (2013) Method of continuous variations: applications of job plots to the study of molecular associations in organometallic chemistry, *Angewandte Chemie International Edition*, **52**: 11998-12013.
31. J. V. Caso, L. Russo, M. Palmieri, G. Malgieri, S. Galdiero, A. Falanga, C. Isernia, R. Iacovino, (2015) Investigating the inclusion properties of aromatic amino acids complexing beta-cyclodextrins in model peptides, *Amino Acids*, **47**: 2215-2227.

32. H. A. Benesi, J. H. Hildebrand, (1949) A spectrophotometric investigation of the interaction of iodine with aromatic hydrocarbons, *Journal of the American Chemical Society*, **71**: 2703-2707.
33. J. V. Caso, L. Russo, M. Palmieri, G. Maltieri, S. Galdiero, A. Falanga, C. Isernia, R. Iacovino, (2015) Investigating the inclusion properties of aromatic amino acids complexing beta-cyclodextrins in model peptides, *Amino Acids*, **47**: 2215-2227.
34. V. Sindelar, M. A. Cejas, F. M. Raymo, W. Chen, S. E. Parker, A. E. Kaifer, (2005) Supramolecular assembly of 2,7-dimethyldiazapyrenium and cucurbit[8]uril: A new fluorescent host for detection of catechol and dopamine, *Chemistry: A European Journal*, **11**: 7054-7059.
35. T. Wang, M. D. Wang, C. Ding, J. Fu, (2014) Mono-benzimidazole functionalized β -cyclodextrins as supramolecular nanovalves for pH-triggered release of p-coumaric acid, *Chemical Communications*, **50**: 12469-12472.
36. E. Bednarek, W. Bocian, J. Poznański, J. Sitkowski, N. S. Sosnowska, L. Kozerski, (2002) Complexation of steroid hormones: prednisolone, ethinylestradiol and estriol with β -cyclodextrin. An aqueous ^1H NMR study, *Journal of Chemical Society Perkin Transactions*, **2**: 999-1004.
37. T. Stalin, K. Srinivasan, K. Sivakumar, S. Radhakrishnan, (2014) Preparation and characterizations of solid/aqueous phases inclusion complex of 2,4-dinitroaniline with β -cyclodextrin, *Carbohydrate Polymers*, **107**: 72-84.
38. B. Rajbanshi, S. Saha, M. N. Roy, (2018) Study to probe subsistence of host-guest inclusion complexes of α and β -cyclodextrins with biologically potent drugs for safety regulatory discharge, *Scientific Reports*, **8**: 13031.
39. N. Roy, R. Ghosh, M. N. Roy, (2019) Study to synthesize and characterize host-guest encapsulation of antidiabetic drug (TgC) and hydroxy propyl- β -cyclodextrin augmenting the antidiabetic applicability in biological system, *Journal of Molecular Structure*, **1179**: 642-650.
40. T. Y. F. Chen, T. C. Chen, F. Y. Cheng, A. T. Tsai, M. T. Lin, (2018) Digital image correlation of SEM images for surface deformation of CMOS, *Microelectronic Engineering*, **201**: 16-21.
41. R. Cossio, C. Albonico, A. Zanella, S. Fraterrigo-Garofalo, C. Avataneo, R. Compagnoni, F. Turci, (2018) Innovative unattended SEM-EDS analysis for asbestos fiber quantification, *Talanta*, **190**: 158-166.

*Bibliographical Sketch



Sri Habibur Rahaman is an Assistant Professor in Chemistry of Pedong Government College, Pedong, Kalimpong, India. He obtained M.Sc. in Chemistry from the University of North Bengal in the year of 2008 and qualified NET (CSIR) and GATE. At present, he has been doing his PhD work in the field of inclusion complexation under the supervision of Prof. Mahendra Nath Roy, Department of Chemistry, NBU. He has done a voluminous work on some significant biologically potent molecules having far-reaching effects in medicinal Chemistry including pharmaceutical and cosmetic products associated with his PhD work and he is going to submit his PhD thesis maintaining official formalities. He has already published four research articles in reputed national and international journals and attended many national and international seminars and conferences for highlighting his research work.



Dr. Mahendra Nath Roy is a senior Professor in the Department of Chemistry, University of North Bengal, India. His research interests are in the areas of host-guest inclusion complexes, ionic liquids, solution thermodynamics, surfactant, polymer chemistry, etc. Prof. Roy has singly produced 40 PhDs, reviewed 37 PhD theses and many referred research papers and authored over 324 research articles and books in Chemistry. He has received the "award of One Time Grant under basic scientific research" from University Grants Commission, "Prof. Suresh C. Ameta Award" from Indian Chemical Society, CRSI Bronze Medal 2017, from Chemical Research Society of India and Shiksha Ratna Award from the Government of West Bengal for his Outstanding Contribution and Excellencies in chemical research throughout his career.

SUPPLEMENTARY FIGURE

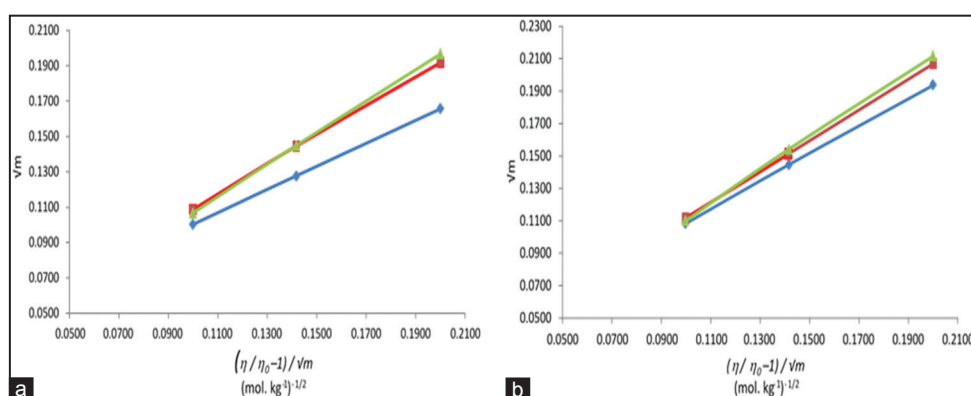


Figure S1: (a) Variation of $(\eta/\eta_0-1)/\sqrt{m}$ against \sqrt{m} of phenylethanolamine in aqueous β -cyclodextrins (b) Variation of $(\eta/\eta_0-1)/\sqrt{m}$ against \sqrt{m} of phenylethanolamine in aqueous α -cyclodextrins. (Blue line for 0.01 m, red line for 0.03 m and green line for 0.05 m cyclodextrins).

SUPPLEMENTARY TABLE

Table S1: Experimental values of density (ρ) and viscosity (η) of different molarities of phenylethanolamine in aqueous α - and β -cyclodextrins at 298.15 K.

Molarity of α -CD in mol/kg	$\rho \times 10^{-3}$ (kg/m ³)	η (mP/s)	Molarity of β -CD in mol/kg	$\rho \times 10^{-3}$ (kg/m ³)	η (mP/s)
Molarity of phenylethanolamine=0.001			Molarity of phenylethanolamine=0.001		
0.010	0.99910	0.9065	0.001	0.99923	0.8204
0.020	0.99956	0.9137	0.002	0.99970	0.8282
0.030	1.00006	0.9273	0.004	1.00018	0.8431
Molarity of phenylethanolamine=0.003			Molarity of phenylethanolamine=0.003		
0.001	0.99933	0.9207	0.001	0.99997	0.8317
0.002	0.99978	0.9294	0.002	1.00043	0.8401
0.004	1.00029	0.9458	0.004	1.00091	0.8566
Molarity of phenylethanolamine=0.005			Molarity of phenylethanolamine=0.005		
0.001	0.99979	0.9282	0.001	0.99931	0.8440
0.002	1.00024	0.9372	0.002	0.99978	0.8530
0.004	1.00074	0.9546	0.004	1.00027	0.8702

Table S2: Experimental values of density (ρ) and viscosity (η) of phenylethanolamine in different molarities of aqueous α - and β -cyclodextrin at 298.15 K.

Molarity (mol/kg)	$\phi_V \times 10^6$ (m ³ /mol ¹)	$(\eta/\eta_0-1)/\sqrt{m}$ (mol/kg) ^{-1/2}	Molarity mol/kg)	$\phi_V \times 10^6$ (m ³ /mol)	$(\eta/\eta_0-1)/\sqrt{m}$ (mol/kg) ^{-1/2}
Molarity of phenylethanolamine=0.001			Molarity of phenylethanolamine=0.001		
0.001	196.5612	0.642	0.001	196.5100	0.645
0.002	195.6088	0.703	0.002	195.3574	0.701
0.004	194.8903	0.782	0.004	194.5054	0.786
Molarity of phenylethanolamine=0.003			Molarity of phenylethanolamine=0.001		
0.001	196.4372	0.647	0.001	196.4214	0.651
0.002	195.2849	0.708	0.002	195.1190	0.712
0.004	194.4333	0.793	0.004	194.0838	0.801
Molarity of phenylethanolamine=0.005			Molarity of phenylethanolamine=0.001		
0.001	196.3978	0.6502	0.001	196.2818	0.655
0.002	195.1457	0.7133	0.002	194.7801	0.715
0.004	194.0605	0.8011	0.004	193.6121	0.809

Table S3: Apparent molar volume (ϕ_V) and $(\eta_r-1)/\sqrt{m}$ of the phenylethanolamine in different molarities of aqueous α - and β -CD mixtures at 298.15 K.

Molarity of phenylethanolamine	$\phi_V^0 \times 10^6$ (m ³ /mol)	$S_V^* \times 10^6$ (m ³ /mol ^{-3/2} .kg ^{1/2})	B (kg ^{1/2} /mol ^{-1/2})	A (kg/mol)
α -CD				
0.001	102.5	-63.46	0.850	0.023
0.003	104.4	-72.09	0.949	0.017
0.005	105.6	78.98	1.01	0.009
β -CD				
0.001	98.19	-41.02	0.653	0.035
0.003	99.60	-45.56	0.828	0.026
0.005	101.4	-54.79	0.898	0.017

CD: Cyclodextrin